

International Journal of Molecular and Immuno Oncology

Review Article

Mini review: Molecular pathology of personalized medicine in cancer susceptibility syndromes

Amrit Kaur Kaler¹, Nandini Shyamali Bora¹, P. Kavyashree¹, Ankita Nikam¹, Samrudhi Rane¹, Yash Tiwarekar¹, Shweta Limaye¹, Varsha Vadera², Mandar Nadkarni³, Yogesh Kulkarni³, T. B. Yuvaraja³, Imran Nisar Shaikh³, Sandeep Goyle³, Rajesh Mistry²

¹Department of Genetics and Molecular Medicine, ²Department of Laboratory Medicine and Advanced Diagnostics, ³Center of Cancer, Kokilaben Dhirubhai Ambani Hospital and Medical Research Institute, Mumbai, Maharashtra, India.



***Corresponding author:**

Amrit Kaur Kaler,
Consultant, Department
of Genetics and Molecular
Medicine, Kokilaben Dhirubhai
Ambani Hospital and
Medical Research Institute,
Maharashtra, Mumbai, India.

amrit_kaler@yahoo.co.in

Received: 14 July 2023
Accepted: 01 August 2023
Epub Ahead of Print: 26 September 2023
Published: 07 October 2023

DOI
10.25259/IJMIO_12_2023

Quick Response Code:



ABSTRACT

In current times, medical oncology is increasingly incorporating cancer genetics and genetic testing into its practice. About 5–10% of all cancers are caused due to inherited genetic mutation that increases susceptibility to a particular malignancy. There is an increasing practice of incorporation of genetic testing and results with potential benefits that have been seen in current-day oncology practice. The American College of Medical Genetics and Genomics highly advises conducting clinical molecular genetic testing within a laboratory that has received CLIA approval with results accurately interpreted by molecular geneticists. The patient is highly recommended to talk to a genetic specialist to explain about the risk, document the family history, and also explain the limitations and outcomes of the genetic testing. Nonetheless, significant discussions and ambiguity persist regarding the optimal approach for providing genetic testing services. These include considerations such as which tests should be employed, which patients should undergo testing, the order and timing of the tests, who should administer them, and the appropriate course of action for follow-up.

Keywords: Molecular pathology, Personalized medicine, Cancer susceptibility syndromes, Solid tumors, Hematological malignancies

INTRODUCTION

Familial cancers can occur in multiple individuals within the same family but are not caused due to single gene mutation.^[1,2] These type of cancers cluster within families but are not hereditary in nature and result from a combination of various factors, such as multiple genes, and lifestyle factors such as diet and exercise, which collectively increases the risk of developing cancers.^[3] Hereditary cancers are caused due to germline mutations in specific genes that are inherited from either one or both parents and are associated with susceptibility to particular cancers. Mostly autosomal dominant in nature, meaning the probability of 50% passing on this mutation or change to the next generation.^[4] An individual who tested positive on a hereditary cancer test has more than one pathogenic variant in 3.1% of cases.^[5] While germline mutations occur in all the cells of the body, cancers that are non-hereditary or sporadic are caused due to genetic mutations in the tumor cells or tissues concerned. These mutations are known as somatic mutations and are not inherited by the next generation.

A few of the cancers follow autosomal recessive patterns of inheritance, such as *MUTYH*-associated polyposis (MAP). MAP is associated with germline mutations in both the copies of

the *MUTYH* gene, one copy of each mutated allele inherited from the maternal and paternal sides. Patients developing colon cancer at an early age and having healthy parents and/or the presence of 15–20 colonic adenomas are indications of the possibility of MAP in an individual.^[6]

Hereditary basis of breast cancer was first described by Broca more than a century ago in 1866 when he reported breast cancer in multiple family members.^[7] Warthin and Henry Lynch reported distinct patterns of early age onset and multiple primary tumors in the same individual with colon cancer.^[8,9] Li and Fraumeni described the clinical characteristics of families with p53 germline mutations as Li-Fraumeni syndrome (LFS) in the subsequent years.^[10] In 1971 Knudson propounded the two-hit hypothesis in Retinoblastoma (RB), where he suggested that the patients who had bilateral RB carried an inherited gene mutation along with an acquired second mutation.^[11] Later, Friend SH *et al.* also confirmed the same hypothesis by and reported a high incidence of 2nd non-ocular tumor which was believed to be caused by the same mutation.^[12]

There have been more than 100 distinct syndromes found, and the majority are rare.^[13] About 5–10% of all cancers are caused due to inherited genetic mutation that increases susceptibility to a particular malignancy.^[4,14] Being familiar with the more-prevalent syndromes, such as hereditary breast and ovarian cancer, Li-Fraumeni, Lynch Syndrome (HPNCC), familial adenomatous polyposis (FAP), RB, multiple endocrine neoplasia, and Von Hippel-Lindau (VHL) can help health-care professionals recognize different signs and symptoms in a patient as potentially having a genetic component, allowing for appropriate diagnostic testing and referrals.^[4]

There has been a tremendous advancement in next-generation sequencing (NGS) technologies in the last 30 years. The application of massively parallel sequencing in germline and somatic cancers has become clinically important. Germline testing assists in identifying the risk of inherited cancer in an individual and at-risk family members and benefit in risk-reducing measures and cancer surveillance.^[14] While somatic testing helps in therapeutic options for targeted therapies and immunotherapies.^[15] Although germline and somatic testing are carried out independently oftentimes in diagnostic laboratories, integrating both approaches to provide optimal care for individuals affected with diverse forms of cancer.^[16-18]

PENETRANCE AND EXPRESSIVITY

Penetrance is a measurement of the proportion of individuals in a population who carry a particular pathogenic mutation and exhibit the disease phenotype. For example, Mutations in the *RBI*, *APC*, *BRCA1* and 2, *PTEN* genes as mentioned in [Table 1]. But in some syndromes, the association between the gene and its expressivity is reduced, like incomplete penetrance shown in Wilms tumor.

The degree to which a genotype manifests its phenotypic expression is measured by expressivity. Different levels of expression in different people may result from variations in the allelic makeup of the rest of the genome or from environmental influences. Thus, expressivity quantifies the degree to which a genotype is phenotypically expressed in individuals, as opposed to penetrance measurements that concentrate on whether or not a disease is expressed in a population.^[19,20]

Understanding the penetrance and expressivity of cancer-predisposing genes is significant in understanding the

Table 1: Few examples of significant cancer susceptibility syndromes, their patterns of inheritance, and penetrance.^[19]

Cancer syndrome	Gene	Main tumor type	Penetrance	Patterns of inheritance
FAP	<i>APC</i>	Colorectal carcinoma	70–100%	Autosomal dominant
Cowden's syndrome	<i>PTEN</i>	Breast, endometrium, follicular thyroid tumor	90–95%	Autosomal dominant
HBOC	<i>BRCA1</i> and <i>BRCA2</i>	Breast/ovary	Up to 85%	Autosomal dominant
LFS	<i>TP53</i>	AML, sarcoma, adrenocortical Carcinoma	90–100%	Autosomal dominant
Lynch syndrome/HNPCC	<i>MLH1</i> , <i>MSH2</i> , <i>MSH6</i> , <i>PMS1</i> , <i>PMS2</i>	Colorectal, endometrium, brain	90%	Autosomal dominant
RB	<i>RB</i>	Eye, bone	90%	Autosomal dominant
Wilms' tumor syndromes	<i>WT1</i>	Nephroblastoma	30% incomplete	Autosomal dominant
Gorlin syndrome/NBCC	<i>PTCH1</i>	Basal cell carcinoma/medulloblastoma	90%	Autosomal dominant
ATS	<i>ATM</i>	Lymphomas, leukemia	100%	Autosomal recessive
FA	<i>FANCA</i> , <i>FANCB</i> , <i>FANCC</i> , <i>FANCD</i> , <i>FANCE</i> , <i>FANCF</i> , <i>FANCG</i> , <i>FANCL</i>	Acute myeloid leukemia	100%	Autosomal recessive
BS	<i>BLM</i>	Wilms tumor, colorectal cancers, Leukemia	100%	Autosomal recessive

FAP: Familial adenomatous polyposis, HBOC: Hereditary breast and ovarian cancer syndrome, LFS: Li-Fraumeni syndrome, HNPCC: Hereditary nonpolyposis colon cancer, NBCC: Nevoid basal cell carcinoma, ATS: Ataxia-telangiectasia syndrome, RB: Retinoblastoma, FA: Fanconi anemia, BS: Bloom syndrome

complexity of hereditary cancers and improving genetic counseling for patients as well as family members.^[19]

INDICATIONS FOR GERMLINE CANCER TESTING INCLUDES

1. Breast cancer diagnosis in ≤50 years age: Triple negative subtype or Lobular Carcinoma; Male Bilateral/multiple primary, Ashkenazi Jewish Ancestry, Breast cancer and one additional tumor (LFS, ≥1 PJ polyp, Cowden syndrome).
2. Colorectal cancer (CRC) diagnosed at age <50: Mismatch repair deficient; multiple primary synchronous or metachronous CRC; ≥10 adenomatous or >5 hamartomatous gastrointestinal polyps; association with other cancers – endometrial, LFS, Cowden syndrome criteria.
3. All women are diagnosed with ovarian cancer whether it is a single case present in the patient or a first-degree relative. *BRCA1* and *BRCA2* pathogenic germline variants are detected in the vast majority of ovarian cancer patients, specifically with high-grade serous histology.
4. Prostate cancer diagnosed at any age: Intraductal/cirriiform histology, Gleason score ≥7; metastatic, regional (node-positive) or very-high-risk localized prostate; Ashkenazi Jewish ancestry.
5. Pancreatic cancer diagnosed at any age: Intraductal papillary mucinous neoplasm histopathology.
6. Patients diagnosed with renal cancer, having age of diagnosis <50; Bilateral or multifocal tumors; ≥1 close relative renal cell carcinoma (RCC) with clear cell, papillary type 1, papillary type 2, collecting duct, tubulopapillary and Birt-Hogg-Dubé (BHD)-related histology, Fumarate hydratase (FH) associated RCC.
7. Thyroid cancers with Medullary subtype, a cirriiform morular subtype of papillary thyroid cancer Papillary/follicular thyroid cancer, and additional carney complex or Cowden syndrome.
8. Gastric cancers: Diffuse type, signet ring cell type, and mismatch repair deficient.
9. Melanoma: Melanoma and pancreatic cancer/ Astrocytoma in the same person.
10. Lynch syndrome-related cancer (i.e., colorectal, endometrial, gastric, ovarian, pancreatic, ureter and renal pelvic, brain (usually glioblastoma), biliary tract, small intestinal, sebaceous adenoma, sebaceous carcinoma, or keratoacanthoma) and the tumor shows evidence of mismatch repair (MMR) deficiency (either by microsatellite instability or loss of MMR protein expression).^[21]

CATEGORIZATION OF GENETIC SYNDROMES BASED ON MOLECULAR PATHWAYS

The syndromes are supposed to be activate pathways that lead to uncontrolled proliferation, increased angiogenesis, or defective repair.

- Defect in PI3K/AKT/mTOR pathway: This is the most common pathway activated in cancer, which leads to the formation of hamartomas/overgrowth syndromes. Cowden syndrome (*PTEN* gene), Proteus syndrome (*AKT1*), Tuberous sclerosis complex (*TSC1/2*), Von Recklinghausen disease (*NF1* and *NF2*).^[22]
- Defect in the RAS/RAF/MEK/ERK pathway: RASopathies, Von Recklinghausen disease (*NF1* and *NF2*).^[23]
- Defect in angiogenesis: *VHL*
- Defects DNA repair mechanism: Hereditary breast and ovarian cancer syndrome, Lynch syndrome, LFS
- Defect in growth factor regulation: Gorlin syndrome (*PTCH1*)
- Others: FAP (*APC*).

TECHNIQUES

NGS-based methods are used to rapidly sequence known cancer-associated genes for identifying germline mutations at once or identify novel germline variants linked to cancer. NGS platforms allow researchers to sequence millions of DNA fragments in parallel, greatly accelerating the process and reducing the cost per base. The high-throughput data generated through NGS make them particularly valuable for understanding complex diseases, including cancers. Among different cancers, there is significant interest in studying those with a familial predisposition, as they offer opportunities to identify novel genes or gene variants that contribute to cancer development and can be detected at the germline level, thus playing a role in cancer pathogenesis.^[24,25]

Multiplex-Ligation Dependent Probe Amplification is another technique that identifies large deletions/duplications in genes. It combines aspects of both polymerase chain reaction and hybridization techniques to analyze the copy number of specific DNA sequences.^[26] Sanger sequencing chain termination sequencing technology is a method used to determine the nucleotide sequence of DNA.^[27] It can be used for mutation confirmation among at-risk family members, siblings, and next generations. It has its limitations in sequencing a single gene as compared to massively parallel sequencing millions of fragments sequenced in NGS.

REPORTING OF VARIANTS ACCORDING TO AMERICAN COLLEGE OF MEDICAL GENETICS AND GENOMICS (ACMG) GUIDELINES

The ACMG guidelines assist in the evaluation and accurate interpretation of genetic variants and have laid down specific criteria for variant classification and reporting based on their association with genetic and clinical presentation. It outlines a systematic approach to evaluating genetic variants based on multiple lines of evidence, such as population frequency,

functional studies, computational, predictive, segregation data, clinical observations, and multiple databases [Table 2]. The guidelines also define standardized terms and criteria for variant interpretation, including pathogenic, likely

pathogenic, uncertain significance, likely benign, and benign, based on the available evidence. The collective evidence from all the criteria evaluated, is then used to assign the appropriate variant classification.^[28]

Table 2: American college of medical genetics and genomics evidences for classifying gene variants.

	Strong	Supporting	Supporting	Moderate	Strong	Very strong
Population data	MAF is too high for disorder BA1/BS1 OR observation in controls inconsistent with disease penetrance BS2			Absent in population databases PM2	Prevalence in affected statistically increased over controls PS4	
Computational and predictive data		Multiple lines of computational evidence suggest no impact on gene/gene product BP4 Missense in gene where only truncating cause disease BP1 Silent variant with non predicted splice impact BP7	Multiple lines of computational evidence support a deleterious effect on the gene/gene product PP3	Novel missense change at an amino acid residue where a different pathogenic missense change has been seen before PM5 Protein length changing variant PM4	Same amino acid change as an established pathogenic variant PS1	Predicted null variant in a gene where LOF is a known mechanism of disease PVS1
Functional data	Well-established functional studies show no deleterious effect BS3		Missense in gene with low rate of benign missense variants and path. Missenses common PP2	Mutational hotspot or well-studied functional domain without benign variation PM1	Well-established functional studies show a deleterious effect PS3	
Segregation data	Non-segregation with disease BS4		N≤1/8 if 1 family N≤1/4 if >1 family	N≤1/16 if 1 family N≤1/8 if >1 family	N≤1/32 if 1 family N≤1/16 if >1 family	
De novo data				De novo (without paternity and maternity confirmed) PM6	De novo (paternity and maternity confirmed) PS2	
Allelic data		Observed in trans with a dominant variant BP2 Observed in cis with a pathogenic variant BP2		For recessive disorders, detected in trans with a pathogenic variant PM3		
Other database		Reputable source without shared data=benign BP6	Reputable source=pathogenic PP5			
Other data		Found in case with an alternate cause BP5	Patient's phenotype or FH highly specific for gene PP4			

GENETIC COUNSELING

Professional societies' consensus statements recommend pre-test and post-test genetic counseling to patients for Germline hereditary cancer genetic testing.^[29] Taking into account the clinical and family history of the patient, informing the patient about indications of genetic testing, as well as addressing the patient's concerns is an important aspect to consider before ordering a genetic test.^[30,31] Constructing a pedigree chart for 3 generations is the best way to visualize risk assessment among patients.^[32,33]

The process of obtaining consent for germline testing should comprise an explanation of the test's rationale, potential outcomes, risks, and advantages. The results of a genetic test should be disclosed followed up with proper post-test genetic counseling.^[31]

Post-test genetic counseling involves providing patients and their families with an in-depth summary of the genetic testing results, helping them to comprehend the test outcomes and associated risks; co-ordinating comprehensive follow-up care to plan for cancer prevention, timely surveillance, and

Table 3: Personalized therapy approvals by FDA and evidence-based studies in germline hereditary cancers.

Gene	Drug	Relationship	Study method
TSC1/2 (mTOR Pathway)	Estrogen-based medications, including oral contraceptives Everolimus, mTOR inhibitor	Shared risk outcome of progression to lymphangioleiomyomatosis TSC-associated Subependymal giant Astrocytoma and renal angiomyolipoma	Case Report ^[36] Case series ^[37] FDA Approved 2018 ^[38]
VHL (Hypoxia induced factor-a)	WELIREG, HIF-2a inhibitors	Sensitivity VHL - RCC, CNS Hemangioblastomas, or pancreatic neuroendocrine tumors, not requiring immediate surgery.	FDA approval in 2021 ^[39]
BRCA1/2 (HRR Pathway)	Oral Contraceptives Clomiphene Citrate Carboplatin, Cisplatin PARP inhibitors	Increased risk of breast cancer (Clinical context) Complete pathological response Increased sensitivity Disease free survival	Retrospective study ^[40] Case-control questionnaire ^[41] Hahnen <i>et al.</i> 2017 ^[42] Caramelo <i>et al.</i> 2019 ^[43] FDA 2018 (Breast), 2020 (Ovary), 2023 (Prostate) ^[44-46] 10-years follow-up of randomized controlled trial ^[47] Phase 1 clinical trial ^[48]
MLH1 MSH2 MSH6 PMS2 TP53	Aspirin and NSAIDs Genotoxic agents like etoposide and radiotherapy Carboplatin and Breast Cancer Aspirin and NSAIDs	Sensitivity Resistance Sensitivity Shared risk outcome	 Review ^[49] <i>In vivo</i> models ^[50] Cohort Study ^[51] FDA approved in 2018 ^[52]
APC (WNT Pathway) RET	Tyrosine Kinase Inhibitors Pralsetinib and selpercatinib	Sensitivity RET mutation-positive medullary thyroid cancer (MEN syndrome)	FDA approved in 2010 ^[53]
SDHA	Tyrosine kinase inhibitors	Resistance (GISTs) Sensitivity (Metastatic PGG and PCC)	Observational study ^[54] Phase 2 clinical trial ^[55]
SDHB	Temozolomide Tyrosine Kinase Inhibitors	Sensitivity Resistance (GISTs)	Retrospective population study ^[56] Observational study ^[54] Case report ^[57] Phase 2 clinical trial ^[55]
SDHC	Tyrosine kinase inhibitors	Sensitivity (Metastatic PGG and PCC) Sensitivity (RCC)	Case Report ^[58]

MEN: Multiple endocrine neoplasia, VHL: Von hippel-lindau, TSC: Tuberous-sclerosis-complex, CNS: Central nervous system, FDA: Food and Drug Administration, NSAIDs: Non-steroidal anti-inflammatory drugs, MEN: Multiple endocrine neoplasia, GIST: Gastrointestinal stromal tumor, RCC: Renal cell carcinoma, PGC: Paraganglioma, PCC: Pheochromocytoma

offering individuals personalized treatment strategies. If an individual is tested “positive” for a germline variant associated with hereditary cancer, other at-risk family members should be encouraged to follow up for genetic counseling.^[34]

MANAGEMENT

Personalized approaches to cancer genetic syndrome

Individuals who test positive for cancer-associated germline mutations require a comprehensive approach to cancer management. Compared to the general population, individuals with cancer germline mutations may necessitate preventive and specialized screening options tailored to the specific associated cancer risks. In certain cases [Table 3], risk reduction surgery options may be recommended to minimize the likelihood of cancer development. For instance, patients consider prophylactic surgeries like mastectomy/oophorectomy/colectomy to decrease the risk of respective cancer.^[35]

Genetic risk predictions models

It has become increasingly common to use computational models in genetic risk prediction models in recent years. A number of risk assessment tools and models are available to evaluate the probability that an individual carries a genetic mutation or their risk to develop cancer. These tools assess the risk based on the presence or absence of gene mutations, personal or family history of cancer. BRCAPRO is an important risk model, developed based on the statistical R package, BayesMendel.^[59] It calculates the individual's probability of carrying a pathogenic *BRCA1* or *BRCA2* gene mutation, the risk of developing contralateral breast cancer and ovarian cancer at different ages. Users can input clinical information such as age, tumor marker information, mastectomy, and oophorectomy information, family race, and ethnicity. BRCAPRO serves as a helpful tool for guiding individuals on whether to pursue genetic testing.^[60]

Another breast cancer risk assessment tool is the Breast Cancer Risk Assessment Tool: Gail Model, named after Gail *et al.*, is a statistical model that uses clinical information such as age, menstruation age, age at first live childbirth, and family history of cancer to estimate the risk of developing cancer.^[61,62]

While these models may help in understanding the risk of an individual, this alone should not be used as a deciding factor to undergo genetic testing as they have certain limitations. Use of the tools might support decisions for a requirement for genetic testing, but it could also lead to stress and anxiety for patients and their family members if the patient is not guided and explained about the risk-assessment model and its implications.

CONCLUSION

Genetic testing for cancer may help in the estimation of an individual's lifetime risk of developing cancer by identifying specific genetic changes or mutations. Germline testing is a powerful tool for early detection and cancer prevention not only in proband cases but also in family members. Understanding the indications for germline testing is a responsibility for coordinating this care between the patient and clinician. Choosing the right test with the right technology will help in the correct interpretation of the results and guide the patient and their family in disease prognosis and awareness of preventative screening options, if available.

Acknowledgment

We thank all the clinicians for their participation in this study and all co-workers in the laboratory for their excellent technical assistance.

Declaration of patient consent

Patient's consent not required as there are no patients in this study.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

Use of artificial intelligence (AI)-assisted technology for manuscript preparation

The author(s) confirms that there was no use of Artificial Intelligence (AI)-Assisted Technology for assisting in the writing or editing of the manuscript and no images were manipulated using the AI.

REFERENCES

1. Frank C, Fallah M, Sundquist J, Hemminki A, Hemminki K. Population landscape of familial cancer. *Sci Rep* 2015;5:12891.
2. Hemminki K, Sundquist K, Sundquist J, Försti A, Hemminki A, Li X. Familial risks and proportions describing population landscape of familial cancer. *Cancers (Basel)* 2021;13:4385.
3. Frank C, Fallah M, Ji J, Sundquist J, Hemminki K. The population impact of familial cancer, a major cause of cancer. *Int J Cancer* 2014;134:1899-906.
4. Rahner N, Steinke V. Hereditary cancer syndromes. *Dtsch Arztebl Int* 2008;105:706-14.
5. Neben CL, Zimmer AD, Stedden W, van den Akker J, O'Connor R, Chan RC, *et al.* Multi-gene panel testing of 23,179 individuals for hereditary cancer risk identifies pathogenic

- variant carriers missed by current genetic testing guidelines. *J Mol Diagn* 2019;21:646-57.
6. Sieber OM, Lipton L, Crabtree M, Heinimann K, Fidalgo P, Phillips RK, *et al.* Multiple colorectal adenomas, classic adenomatous polyposis, and germ-line mutations in MYH. *N Engl J Med* 2003;348:791-9.
 7. Broca P. *Traite des Tumeurs*. Paris: P. Asselin; 1866.
 8. Warthin AS. Heredity with reference to carcinoma as shown by the study of the cases examined in the Pathological Laboratory of the University of Michigan, 1895-1912. *Arch Int Med* 1913;12:546-55.
 9. Lynch HT, Shaw MW, Magnuson CW, Larsen AL, Krush AJ. Hereditary factors in cancer. Study of two large midwestern kindreds. *Arch Intern Med* 1966;117:206-12.
 10. Li FP, Fraumeni JF Jr. Soft-tissue sarcomas, breast cancer, and other neoplasms. A familial syndrome? *Ann Intern Med* 1969;71:747-52.
 11. Knudson AG Jr. Mutation and cancer: Statistical study of retinoblastoma. *Proc Natl Acad Sci U S A* 1971;68:820-3.
 12. Friend SH, Bernards R, Rogelj S, Weinberg RA, Rapaport JM, Albert DM, *et al.* A human DNA segment with properties of the gene that predisposes to retinoblastoma and osteosarcoma. *Nature* 1986;323:643-6.
 13. Rahman N. Realizing the promise of cancer predisposition genes. *Nature* 2014;505:302-8.
 14. Garber JE, Offit K. Hereditary cancer predisposition syndromes. *J Clin Oncol* 2005;23:276-92.
 15. Chakravarty D, Johnson A, Sklar J, Lindeman NI, Moore K, Ganesan S, *et al.* Somatic genomic testing in patients with metastatic or advanced cancer: ASCO provisional clinical opinion. *J Clin Oncol* 2022;40:1231-58.
 16. Hicks JK, Howard R, Reisman P, Adashek JJ, Fields KK, Gray JE, *et al.* Integrating somatic and germline next-generation sequencing into routine clinical oncology practice. *JCO Precis Oncol* 2021;5:884-95.
 17. Yap TA, Stadler ZK, Stout LA, Schneider BP. Aligning germline cancer predisposition with tumor-based next-generation sequencing for modern oncology diagnosis, interception, and therapeutic development. *Am Soc Clin Oncol Educ Book* 2023;43:e390738.
 18. Liu YL, Stadler ZK. The future of parallel tumor and germline genetic testing: Is there a role for all patients with cancer? *J Natl Compr Canc Netw* 2021;19:871-8.
 19. Taeubner J, Wiczorek D, Yasin L, Brozou T, Borkhardt A, Kuhlen M. Penetrance and expressivity in inherited cancer predisposing syndromes. *Trends Cancer* 2018;4:718-28.
 20. Kingdom R, Wright CF. Incomplete penetrance and variable expressivity: From clinical studies to population cohorts. *Front Genet* 2022;13:920390.
 21. Stoffel EM, Carethers JM. Current approaches to germline cancer genetic testing. *Annu Rev Med* 2020;71:85-102.
 22. Piombino C, Cortesi L, Lambertini M, Punie K, Grandi G, Toss A. Secondary prevention in hereditary breast and/or ovarian cancer syndromes other than BRCA. *J Oncol* 2020;2020:6384190.
 23. Tamura R. Current understanding of neurofibromatosis Type 1, 2, and schwannomatosis. *Int J Mol Sci* 2021;22:5850.
 24. Price KS, Svenson A, King E, Ready K, Lizarin GA. Inherited cancer in the age of next-generation sequencing. *Biol Res Nurs* 2018;20:192-204.
 25. Zelli V, Compagnoni C, Cannita K, Capelli R, Capalbo C, Di Vito Nolfi M, *et al.* Applications of next generation sequencing to the analysis of familial breast/ovarian cancer. *High Throughput* 2020;9:1.
 26. Stuppia L, Antonucci I, Palka G, Gatta V. Use of the MLPA assay in the molecular diagnosis of gene copy number alterations in human genetic diseases. *Int J Mol Sci* 2012;13:3245-76.
 27. Sanger F, Nicklen S, Coulson AR. DNA sequencing with chain-terminating inhibitors. *Proc Natl Acad Sci U S A* 1977;74:5463-7.
 28. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, *et al.* Standards and guidelines for the interpretation of sequence variants: A joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015;17:405-24.
 29. Bokkers K, Bleiker EM, Velthuis ME, Koelemij R, Burgmans JP, Klinkenbijn JH, *et al.* Patients' experiences with pre-test genetic counseling provided by breast cancer healthcare professionals: Results from a large prospective multicenter study. *Breast* 2023;69:349-57.
 30. Institute of Medicine (US) Committee on Assessing Genetic Risks, Andrews LB, Fullerton JE, Holtzman NA, Motulsky AG, editors. *Assessing Genetic Risks: Implications for Health and Social Policy*. Washington, DC: National Academies Press (US); 1994.
 31. Bennett RL, Hampel HL, Mandell JB, Marks JH. Genetic counselors: Translating genomic science into clinical practice. *J Clin Invest* 2003;112:1274-9.
 32. Mahon SM. The three-generation pedigree: A critical tool in cancer genetics care. *Oncol Nurs Forum* 2016;43:655-60.
 33. Bennett RL, Steinhaus KA, Uhrich SB, O'Sullivan CK, Resta RG, Lochner-Doyle D, *et al.* Recommendations for standardized human pedigree nomenclature. Pedigree Standardization Task Force of the National Society of Genetic Counselors. *Am J Hum Genet* 1995;56:745-52.
 34. American Society of Human Genetics Board of Directors, American College of Medical Genetics Board of Directors. Points to consider: Ethical, legal, and psychosocial implications of genetic testing in children and adolescents. American Society of Human Genetics Board of Directors, American College of Medical Genetics Board of Directors. *Am J Hum Genet* 1995;57:1233-41.
 35. Saugstad AA, Petry N, Hajek C. Pharmacogenetic review: Germline genetic variants possessing increased cancer risk with clinically actionable therapeutic relationships. *Front Genet* 2022;13:857120.
 36. Yano S. Exacerbation of pulmonary lymphangiomyomatosis by exogenous oestrogen used for infertility treatment. *Thorax* 2002;57:1085-6.
 37. Oberstein EM, Fleming LE, Gómez-Marin O, Glassberg MK. Pulmonary lymphangiomyomatosis (LAM): Examining oral contraceptive pills and the onset of disease. *J Womens Health (Larchmt)* 2003;12:81-5.
 38. Available from: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-everolimus-tuberous-sclerosis-complex-associated-partial-onset-seizures> [Last accessed on 23 Jul 14].
 39. Available from: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-belzutifan-cancers-associated-von>

- hippel-lindau-disease [Last accessed on 23 Jul 14].
40. Huber D, Seitz S, Kast K, Emons G, Ortmann O. Use of oral contraceptives in BRCA mutation carriers and risk for ovarian and breast cancer: A systematic review. *Arch Gynecol Obstet* 2020;301:875-84.
 41. Reigstad MM, Storeng R, Myklebust Å TÅ, Oldereid NB, Omland AK, Robsahm TE, *et al.* Cancer risk in women treated with fertility drugs according to parity status-a registry-based cohort study. *Cancer Epidemiol Biomarkers Prev* 2017;26:953-62.
 42. Hahnen E, Lederer B, Hauke J, Loibl S, Kröber S, Schneeweiss A, *et al.* Germline mutation status, pathological complete response, and disease-free survival in triple-negative breast cancer: Secondary Analysis of the GeparSixto Randomized Clinical Trial. *JAMA Oncol* 2017;3:1378-85.
 43. Caramelo O, Silva C, Caramelo F, Frutuoso C, Almeida-Santos T. The effect of neoadjuvant platinum-based chemotherapy in BRCA mutated triple negative breast cancers-systematic review and meta-analysis. *Hered Cancer Clin Pract* 2019;17:11.
 44. Available from: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-olaparib-germline-brca-mutated-metastatic-breast-cancer> [Last accessed on 23 Jul 14].
 45. Available from: <https://www.fda.gov/drugs/resources-information-approved-drugs/fda-approves-olaparib-plus-bevacizumab-maintenance-treatment-ovarian-fallopian-tube-or-primary> [Last accessed on 23 Jul 14].
 46. Available from: <https://www.fda.gov/drugs/drug-approvals-and-databases/fda-approves-olaparib-abiraterone-and-prednisone-or-prednisolone-brca-mutated-metastatic-castration>
 47. Burn J, Sheth H, Elliot F, Reed L, Macrae F, Mecklin JP, *et al.* Cancer prevention with aspirin in hereditary colorectal cancer (lynch syndrome), 10-year follow-up and registry-based 20-year data in the CAPP2 study: A double-blind, randomised, placebo-controlled trial. *Lancet* 2020;395:1855-63.
 48. Reyes-Urbe L, Wu W, Gelinck O, Bommi PV, Francisco-Cruz A, Solis LM, *et al.* Naproxen chemoprevention promotes immune activation in Lynch syndrome colorectal mucosa. *Gut* 2021;70:555-66.
 49. Frebourg T, Lagercrantz SB, Oliveira C, Magenheim R, Evans DG, European Reference Network GENTURIS. Guidelines for the Li-Fraumeni and heritable TP53-related cancer syndromes. *Eur J Hum Genet* 2020;28:1379-86.
 50. Kasper E, Angot E, Colasse E, Nicol L, Sabourin JC, Adriouch S, *et al.* Contribution of genotoxic anticancer treatments to the development of multiple primary tumours in the context of germline TP53 mutations. *Eur J Cancer* 2018;101:254-62.
 51. Sheng S, Xu Y, Guo Y, Yao L, Hu L, Ouyang T, *et al.* Prevalence and clinical impact of TP53 germline mutations in Chinese women with breast cancer. *Int J Cancer* 2020;146:487-95.
 52. FDA. CELEBREX® (Celecoxib). Silver Spring, MD: U.S Food and Drug Administration; 2018.
 53. FDA. VICTOZA® (Liraglutide) Injection, for Subcutaneous Use. Silver Spring, MD: U.S Food and Drug Administration; 2010.
 54. Boikos SA, Pappo AS, Killian JK, LaQuaglia MP, Weldon CB, George S, *et al.* Molecular subtypes of KIT/PDGFRα wild-type gastrointestinal stromal tumors: A report from the national institutes of health gastrointestinal stromal tumor clinic. *JAMA Oncol* 2016;2:922-8.
 55. O’Kane GM, Ezzat S, Joshua AM, Bourdeau I, Leibowitz-Amit R, Olney HJ, *et al.* A phase 2 trial of sunitinib in patients with progressive paraganglioma or pheochromocytoma: The SNIPP trial. *Br J Cancer* 2019;120:1113-9.
 56. Hadoux J, Favier J, Scoazec JY, Leboulleux S, Al Ghuzlan A, Caramella C, *et al.* SDHB mutations are associated with response to temozolomide in patients with metastatic pheochromocytoma or paraganglioma. *Int J Cancer* 2014;135:2711-20.
 57. Paik JY, Toon CW, Benn DE, High H, Hasovitz C, Pavlakis N, *et al.* Renal carcinoma associated with succinate dehydrogenase B mutation: A new and unique subtype of renal carcinoma. *J Clin Oncol* 2014;32:e10-3.
 58. Shuch B, Agochukwu N, Ricketts CJ, Vocke CD, Gautam R, Merino M, *et al.* Vascular endothelial growth factor receptor-targeted therapy in succinate dehydrogenase C kidney cancer. *J Clin Oncol* 2016;34:e76-9.
 59. Chen S, Wang W, Broman KW, Katki HA, Parmigiani G. BayesMendel: An R environment for Mendelian risk prediction. *Stat Appl Genet Mol Biol* 2004;3:21.
 60. Mazzola E, Blackford A, Parmigiani G, Biswas S. Recent Enhancements to the Genetic Risk Prediction Model BRCAPRO. *Cancer Inform* 2015;14:147-57.
 61. Gail MH, Brinton LA, Byar DP, Corle DK, Green SB, Schairer C, *et al.* Projecting individualized probabilities of developing breast cancer for white females who are being examined annually. *J Natl Cancer Inst* 1989;81:1879-86.
 62. Stevanato KP, Pedroso RB, Dell Agnolo CM, Santos LD, Pelloso FC, Carvalho MD, *et al.* Use and applicability of the gail model to calculate breast cancer risk: A scoping review. *Asian Pac J Cancer Prev* 2022;23:1117-23.

How to cite this article: Kaler AK, Bora NS, Kavyashree P, Nikam A, Rane S, Tiwarekar Y, *et al.* Mini review: Molecular pathology of personalized medicine in cancer susceptibility syndromes. *Int J Mol Immuno Oncol* 2023;8:81-8.